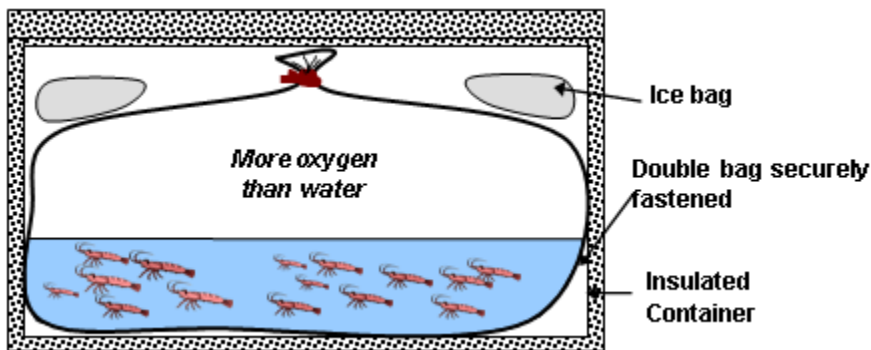


Method of Sampling

Sending of live shrimp samples

1. Obtain 3-5 diseased or moribund shrimps and an equal number of normal shrimps.
2. Pack separately and aerate or place oxygen in plastic bags.
3. Place bag in a styrofoam box or cooler to keep temperature cool.
4. Place small packs of ice when traveling long distances or during hot weather.



NOTE:

For larvae or postlarvae stages, pack at least 175 diseased samples in the same manner.

Sending of fixed shrimp samples for PCR analysis

1. Samples should be fixed in 95% ethanol in a tightly capped bottle or plastic container.
2. For large specimens, dissect the gills and fix in 95% ethanol.
3. Samples should be submitted to the laboratory as soon as possible.

NOTE:

During dissection, scissors, forceps, and scalpels should be disinfected by wiping or dipping in 70% alcohol. Dissecting instruments may be flamed and allowed to cool before use.

Sending of fixed shrimp samples for Histopathology

1. Inject Davidsons fixative (0.1 to 10 ml depending on size of shrimp) using syringe into live shrimp.
2. Insert the needle:

(A) laterally into the hepatopancreas,



(B) in the posterior abdominal segment,



(C) in the region anterior to the hepatopancreas, and



(D) in the anterior abdominal region



3. The incision in the cephalothoracic region should be just lateral to the dorsal midline, while that in the abdominal region should be approximately mid-lateral.
4. Shrimp larger than 12 grams, should be transversely slit once at the:

(E) abdomen/cephalothorax junction and



(F) mid-abdominally



5. Immerse the specimen in the fixative. A rule of thumb is that approximately 10 times the volume of fixative should be used for specimen.